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Note

A new sesquiterpene ester from the fruits of *Celastrus orbiculatus*

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From the fruits of *Celastrus orbiculatus* Thunb. a new β -dihydroagarofuran sesquiterpene ester named 1 β ,2 β ,13-triacetoxy-9 α -cinnamoyloxy- β -dihydroagarofuran (1) has been isolated along with two known compounds, 6α -acetoxy-1 β ,9 β -dibenzoyloxy-8 β -hydroxy- β -dihydroagarofuran (2), and 1 β , 6α -diacetoxy-9 β -benzoyloxy-8 β -hydroxy- β -dihydroagarofuran (3). Their structures were elucidated on the basis of spectroscopic data.

Keywords: Celastrus orbiculatus; Sesquiterpene ester; 1β , 2β ,13-Triacetoxy- 9α -cinnamoyloxy- β -dihydroagarofuran

1. Introduction

Celastrus orbiculatus, a medicinal plant widely distributed in China, has activity in tranquilization [1]. Some sesquiterpenes with anti-inflammatory activities from *Celastrus orbiculatus* have been reported [2]. We reported a new β -dihydroagarofuran sesquiterpene ester in previous paper [3]. In extending our research, a new structural elucidation of the new sesquiterpene ester.

2. Results and discussion

Compound **1** was isolated as a white powder (mp 207–209°C). HRMS gave a molecular ion peak at m/z 542.2520 [M]⁺, corresponding to a molecular formula of C₃₀H₃₈O₉ for **1**. The ¹H NMR spectrum gave proton signals at δ 1.21 (3H, s, H-15), 1.27 (3H, d, J = 8.7 Hz, H-12),

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1.39 (3H, s, H-14), 1.80 (3H, s), 2.10 (3H, s), 2.17 (3H, s). The ¹³C NMR spectrum gave three methyl carbon signals at δ 18.5 (C-12), 24.0 (C-14), 30.0 (C-15), four methylene carbon signals at δ 30.8 (C-3), 36.1 (C-6), 33.8 (C-8), 65.2 (C-13), five methine carbon signals at δ 71.1 (C-1), 70.1 (C-2), 39.2 (C-4), 43.4 (C-7), 69.2 (C-9), and three quaternary carbon signals at δ 86.3 (C-5), 50.4 (C-10), 81.9 (C-11). These spectral data suggest a β -dihydroagarofuran sesquiterpene-type skeleton [4,5].

The ¹³C NMR spectrum gave four carbon signals at δ 71.1 (C-1), 70.1 (C-2), 69.2 (C-9), 65.2 (C-13), indicating the presence of four oxygenated carbons. According to the literatures [4,5], the carbonyl carbon signals at δ 169.5, 169.8, 170.3 in the ¹³C NMR spectrum show that compound **1** contains three acetoxy groups; the aromatic carbon signals at δ 134.2–128.1, the carbonyl carbon signal at δ 165.8 and the two carbon signals at δ 117.9, 145.2 as well as the proton signals at δ 7.68, 6.38 (each 1H, d, J = 15.9 Hz), 7.35–7.55 showed the presence of one cinnamoyloxy group.

In the HMQC spectrum, the correlations of signals seen at δ 5.68 (1H, d, J = 3.0 Hz, H-1) with 71.1(C-1), δ 5.54 (1H, dd, J = 6.6, 3.0 Hz, H-2) with 70.1 (C-2), δ 5.19 (1H, d, J = 6.6 Hz, H-9) with 69.2 (C-9), and δ 4.44, 4.82 (each 1H, d, J = 12.6 Hz, H-13) with 65.2 (C-13) are compatible with above conclusion.

In the HMBC spectrum, the proton signal of H-1 at δ 5.68 (1H, d, J = 3.0 Hz) shows long-range correlations with the carbon signals at δ 70.1 (C-2), 30.8 (C-3), 86.3 (C-5), 50.4 (C-10), 65.2 (C-13) and 169.5. Long-range correlations of signals at δ 5.54 (1H, dd, J = 6.6, 3.0 Hz, H-2) with 71.1 (C-1), 30.8 (C-3), 39.2 (C-4), 50.4 (C-10), 169.8, and δ 4.44, 4.82 (each 1H, d, J = 12.6 Hz, H-13) with 71.1 (C-1), 50.4 (C-10), 69.2 (C-9), 86.3 (C-5), 170.3 were also observed; hence the three acetoxy groups are at C-1, C-2, C-13 respectively. The proton signal at δ 5.19 (1H, d, J = 6.6 Hz, H-9) show long-range correlations with the carbon signals at δ 33.8 (C-8), 43.4 (C-7), 50.4 (C-10), 65.2 (C-13), 71.1 (C-1), 86.3 (C-5), 165.8, therefore one cinnamoyloxy group is linked at C-9.

The configurations of the substituted groups of the dihydroagarofuran skeleton were determined by the coupling constants of protons ($J_{1,2} = 3.0$ Hz, $J_{8,9} = 6.6$ Hz) compared with those reported in previously [4–6]. Thus, compound **1** was identified as 1 β ,2 β ,13-triacetoxy-9 α -cinnamoyloxy- β -dihydroagarofuran (figure 1).

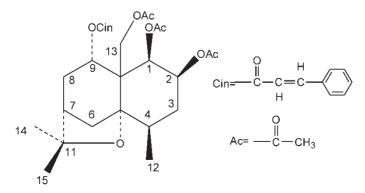


Figure 1. Structure of compound 1.

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3. Experimental

3.1 General experimental procedures

Melting points were measured on a Yamaco-hot-stage and are uncorrected. All NMR spectra were recorded on a Bruker-ARX-300 spectrometer, using TMS as an internal standard. The UV spectrum was recorded on a Shimadzu UV-260 UV–Vis spectrometer. ESI-MS was performed on a VG-70SE mass spectrometer. The optical rotation was measured on a Perkin–Elmer 241 polarimeter. Silica gel for chromatography was produced by the Qingdao Ocean Chemical Group Co. of China. The HPLC system used a Shimadzu CTO-6A equipped with a UV detector, and a Shimadzu SPD-6A column (Shimadzu Shim-pack PREP-ODS, i.d. 2.5×21.6 cm). The boiling point range of light petroleum was $60-90^{\circ}$ C.

3.2 Plant material

The plant material was collected in Shenyang city, Liaoning Province, China, and was identified by Professor Guo Yunzheng (Department of Pharmacognosy, Shenyang Pharmaceutical University, China). A voucher specimen (200115) has been deposited in the Herbarium of the Research Department of Natural Medicine, Shenyang Pharmaceutical University, China.

3.3 Extraction and isolation

Dried fruits (10 kg) of *Celastrus orbiculatus* were extracted with 95% ethanol. The EtOH extract was concentrated *in vacuo*, and then the residue was extracted with light petroleum, EtOAc and n-BuOH successively. A portion (160 g) of the light petroleum extract was subjected to column chromatography on silica gel and eluted with light petroleum–acetone (100:6) to yield compound **1** (5 mg); light petroleum–acetone (100:7) provided fraction 5,

No.	δ_C	δ_H	НМВС
1 ^a	71.1	5.68 (1H, d, $J = 3.0 \text{Hz}$)	C-2, C-3, C-5, C-10, C-13
2 ^a	70.1	5.54 (1H, dd, J = 6.6, 3.0 Hz)	C-1, C-3, C-4, C-10
3	30.8	1.76 (1H, dd, J = 15.1, 1.8 Hz)	C-2, C-5, C-1, C-4
		2.45 (1H, ddd, $J = 15.1, 6.8, 3.4 \text{ Hz}$)	
4	39.2	1.94 (1H, m)	C-12, C-2, C-3, C-5, C-10, C-6
5	86.3		
6	36.1	2.28 (2H, m)	C-4, C-5, C-7, C-8, C-10, C-11
7	43.4	205 (1H, m)	C-6, C-5, C-8, C-9, C-11
8	33.8	2.27 (2H, m)	C-6, C-7, C-10, C-9, C-11
9 ^a	69.2	5.19 (1H, d, J = 6.6 Hz)	C-8, C-7, C-10, C-13, C-1, C-5
10	50.4		
11	81.9		
12	18.5	1.27 (3H, d, $J = 8.7$ Hz)	C-5, C-3, C-4
13*	65.2	4.82 (1H, d, $J = 12.6$ Hz)	C-1, C-10, C-9, C-5
		4.44 (1H, d, $J = 12.6 \text{Hz}$)	
14	24.0	139 (3H, s)	C-15, C-11, C-7
15	30.0	1.27 (3H, s)	C-14, C-11, C-7

Table 1. ¹H and ¹³C NMR data for **1** in CDCl₃.

All signals assigned by 1D and 2D NMR. *Signals of H-1, H-2, H-9 and H-13 were also correlated with carbonyl carbon signals at δ 169.5, 169.8, 165.8, 170.3 respectively.

which was then subjected to HPLC using MeOH $-H_2O(75:25)$ as eluent to yield compounds **2** (10 mg, 40 min) and **3** (11 mg, 57 min).

Compound 1: a white powder (EtOAc), mp 207–209°C; $[\alpha]_D = +75.0$ (c = 0.075 MeOH). UV λ_{max} (MeOH): 279.6 nm. ¹H NMR (300 MHz, in CDCl₃) see table 1, acetate: δ (ppm): 1.80 (3H, s), 2.10 (3H, s), 2.17 (3H, s); cinnamoyloxy: δ 7.68, 6.38 (each 1H, d, J = 15.9 Hz), 7.37–7.55 overlapped. ¹³C NMR (75 MHz, in CDCl₃) see table 1, acetate: δ (ppm): 20.5, 21.1, 21.2, 169.5, 169.8, 170.3; cinnamoyloxy: δ 165.8, 117.9, 145.2, 134.2, 130.2, 128.7, 128.1. EIMS m/z (rel. int. %): 542 [M]⁺ (2) 411 (57) 351 (25) 291 (20) 231 (12) 131 (100) 103 (28) 43 (25); HRMS: m/z 542.2520 [M]⁺ (calcd for C₃₀H₃₈O₉, 542.2516).

¹H and ¹³C NMR data of compounds **2** and **3** are in accordance with the known compounds 6α -acetoxy-1 β ,9 β -dibenzoyloxy-8 β -hydroxy- β -dihydroagarofuran [5] and 1 β , 6α -diacetoxy-9 β -benzoyloxy-8 β -hydroxy- β -dihydroagarofuran [7] respectively.

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